

A step forward for gene editing: CRISPR-Cas9 and HD

Evolving CRISPR-Cas9 techniques can now be used to edit the HD gene in a living mouse brain.



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C *CRISPR-Cas9 is an experimental gene editing technique used to make precise changes in DNA. For the first time, scientists have used this approach to attack the Huntington's Disease mutation in the brain cells of a mouse. Other researchers are refining CRISPR-Cas9 to be more efficient, specific, and safe. It's still a long way from use in HD patients, but its application in mice is an exciting step forward.*

Gene editing with CRISPR-Cas9

DNA is the fundamental code directing the growth and function of living cells. Our ability to manipulate this code, once the stuff of science fiction, has begun to drive research on inherited disorders like Huntington's disease. The design and application of tools to change DNA is known as *gene editing*, and one tool that has received recent attention is called CRISPR-Cas9. Since its introduction, HD scientists have been exploring the possibility that CRISPR-Cas9 could be used to edit out the genetic mutation that causes HD.



Gene editing techniques result in a permanent change to the DNA code

This experimental technique is not ready to try in humans, but it has moved quickly from test tubes to living cells to organisms. Recent work from multiple research groups has shown that CRISPR-Cas9 can be used to edit the HD gene in the brain of a living mouse. Even more exciting, one laboratory's newest findings show improved HD mouse behavior after delivering CRISPR-Cas9 to the brain. This gene editing technology continues to become more sophisticated, and several HD research teams are adapting it to the challenges of HD therapy. Let's talk about how CRISPR-Cas9 works, its application to HD, and why safety is a concern at this stage.

The HD gene: editing the story

HD is caused by an unwanted addition to the genetic code. Billions of biological building blocks, the nucleotides C,G,A and T, are found in the full DNA code. These nucleotides are read and interpreted in chunks - sections of the gene known as exons. You can think of a nucleotide as a letter, three nucleotides as a word, an exon as a sentence, the gene as a paragraph, and the complete genome as an instruction manual describing all the parts needed for cells to grow and function.

Let's zoom in on one paragraph of the story, the gene that causes HD. In people destined to develop HD the first sentence contains a mistake: a string of C-A-G letters that goes on... and on and on and on and on and on and on and on and on and on and on and on and on and on...many times more than necessary. What if we could edit the CAG repeat mistake, like deleting all those extra "on-and-on" repeats from the previous sentence? This is the main focus of gene editing in HD research, and CRISPR-Cas9 is one of several approaches.

CRISPR-Cas9: making the cut

There's no equivalent of word processing software to edit genes. To fix genes on a microscopic scale, one cell at a time, the faulty code has to be located and physically cut – and that's what CRISPR-Cas9 does. This cutting requires two components: (1) a **guide RNA** and (2) a cutting enzyme called **Cas9**. Here's a simple analogy: imagine you want to cut a piece of ribbon, but your friend has the scissors. You might hold the ribbon with two hands, pulling tight to show your friend exactly where to make the cut. That's CRISPR-Cas9, on a microscopic scale: the guide RNA finds and presents the right spot on the DNA, and the Cas9 acts as the scissors, actually cutting the DNA.

In the laboratory, scientists can design specific guide RNAs that will show Cas9 where to cut twice, on both sides of the extra long stretch of C-A-G repeats in the HD gene. Then the new ends can be patched together, permanently removing the offending part. This is how scientists use CRISPR-Cas9 to *edit* genetic sequences.

As with any exciting new technology, researchers have been playing with the CRISPR-Cas9 system to discover novel ways to use the tool. Early on, researchers realized they could use CRISPR-Cas9 to make single cuts in a specific gene pretty easily. The repair process cells use to fix these cuts is error prone, and commonly leads to tiny pieces of the genetic information being lost.

As an analogy, imagine you were typing a text to a friend at the dinner table that said: “please pass the butter”. If you inadvertently skipped a couple letters - say “le” - but kept the structure of the message your friend would receive: “pasepa ssth ebu tterxx”. When genetic messages get jumbled with small deletions like this, cells have machinery for recognizing the errors and ignoring their contents. This gives researchers a way to use CRISPR-Cas9 to effectively *delete* a gene, rather than *editing* the sequence in a more specific way.

CRISPR-Cas9 in an HD mouse brain

A couple of research groups have just discovered that it’s possible to edit the HD gene in the brain of a living mouse. Most recently, a team led by Xiao-Jiang Li, working at Emory University in the USA, found that making tiny cuts in the HD gene could have beneficial effects in HD mice. For these experiments, they were using CRISPR-Cas9 in the *delete* mode, rather than *editing* the HD gene to remove the long C-A-G with a short one.

To use CRISPR-Cas9 in an HD mouse, the guideRNA and Cas9 “scissors” are carried by specially designed viruses that must be injected into the brain. Li’s group applied this technique to the striatum, a brain area controlling mood and movement that becomes damaged during HD. A few weeks later, the CRISPR-cas9 components had spread to many cells, disabling the dysfunctional HD gene, and signs of stress on the neurons had diminished.

After three months, there were fewer harmful clumps of huntingtin protein built up in brain cells, and the HD mice had improved somewhat on movement tests. The most exciting aspect of this experiment was the recovery of older mice that had already developed symptoms. Even 9-month-old mice (around middle age) improved after receiving the injections, suggesting that their brains could recuperate partially after half a lifetime of damage.

Proceeding with caution

Most people with HD have only one copy of the mutant gene, and another copy that is perfectly healthy. There is some concern about using CRISPR-Cas9 as a therapy, because while it could delete the damaged portion from the HD gene, it could also permanently remove a part of the healthy copy. The Li lab also did some experiments to indirectly

address this question, working with mice that had *two* faulty copies of the HD gene, and using CRISPR-Cas9 to remove both. There was no immediate danger to the mice, though they were only monitored for a few weeks.

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The safety of interfering with the normal copy of the HD gene is important, considering the ongoing clinical trial of a huntingtin-lowering ASO. The drug reduces levels of both the mutant and the healthy copy of the HD gene. Some research in mice has suggested that this is harmless during later life, but it's difficult to be sure, because a mouse's lifespan is much shorter than a human's. The companies conducting the ASO study - Roche and Ionis - are well aware of these risks, and are carefully monitoring subjects in the study for any signs of problems caused by lowering the HD gene.

There are other key differences between ASO drugs and the CRISPR-Cas9 approach. The current ASO trial in humans is a *huntingtin lowering*, or *gene silencing* therapy, which works to disable both copies of the HD gene in short bursts. If the treatment is halted, the gene will recover its function. Conversely, *gene editing* using CRISPR-Cas9 creates a permanent change to the DNA, and therefore must be approached with even more caution. There's evidence that the HD gene, damaged or not, has important functions in the cell, and we don't want to risk permanent side effects. The good news is that HD scientists are tackling the challenge of avoiding the healthy copy of the gene, known as an allele-specific approach.

Improving gene editing techniques

Two groups have recently improved upon the CRISPR-Cas9 technique, using it to cut and inactivate only the damaged copy of the gene. A team led by Jong-Min Lee at Massachusetts General Hospital made an allele-specific deletion using cleverly designed guide RNAs. The guides sought out tiny discrepancies in the DNA letters close to the HD mutation and directed two Cas9 cuts. Their approach is novel because gene edits could be “personalized” depending on an individual's DNA.

A second group, headed by Beverly Davidson at the Children's Hospital of Philadelphia, used a similar approach to target only the mutant gene, making smaller cuts with Cas9. This stopped the production of multiple harmful huntingtin proteins. Like Li's group, they could also inactivate the HD gene in the brain of a living mouse. Whether either updated CRISPR technique will improve the behavior of an HD mouse remains to be seen, but both innovations are a step towards the genetic therapies of the future.

Challenges to gene editing

We're excited about the use of gene editing to better understand HD. The use of CRISPR in a living mouse and the development of allele-specific approaches represent significant

steps forward, but there are several hurdles to overcome before CRISPR-Cas9 could be developed into an HD treatment. Here are the main challenges that researchers face, and our current state of knowledge:

1. **Precision:** making sure that the Cas9 is only cutting the gene it is designed to cut, and not doing random harm elsewhere. Scientists seem well on the way to ensuring that CRISPR is very specific.
2. **Allele-specificity:** ensuring that only the mutant copy of the HD gene, and not the healthy one, is removed. The research we have described here is an exciting step forward.
3. **Delivery:** getting the CRISPR-Cas9 machinery to enter many neurons in the brain and remove the HD gene from each. Now we know it's possible in a mouse, but it remains a major hurdle for any therapy used to treat the human brain.
4. **Short-term safety:** making sure that deleting part of the HD gene won't cause immediate neurological problems or even death. So far this appears to be the case.
5. **Long-term safety:** making sure that editing the HD gene would be safe over a long period of time. This is a very difficult question to explore in mice. We may find answers through primate experiments, or from less permanent techniques in clinical trials.

The huntingtin lowering ASO trial is still in early safety stages, but the approach has shown promise so far. Gene editing could introduce enduring changes to what's written in the DNA code, with profound consequences. Safely using CRISPR-Cas9 becomes exponentially more challenging as it moves closer to the clinic. Nevertheless, the next generation of this technology shows incredible promise, and many minds are moving it forward in innovative ways.

Jeff Carroll, editor of this piece, has a long standing, non-financial, research collaboration with Ionis Pharmaceuticals whose ASO trial is discussed in this article. Ed Wild, HDBuzz co-editor-in-chief, is a study investigator in the Ionis/Roche ASO trial, mentioned in the article. Neither Dr. Wild nor employees of Ionis or Roche had input into the writing or editing of this article. [For more information about our disclosure policy see our FAQ...](#)

GLOSSARY

huntingtin protein The protein produced by the HD gene.

gene silencing An approach to treating HD that uses targeted molecules to tell cells not to produce the harmful huntingtin protein

clinical trial Very carefully planned experiments designed to answer specific questions about how a drug affects human beings

CAG repeat The stretch of DNA at the beginning of the HD gene, which contains the sequence CAG repeated many times, and is abnormally long in people who will

develop HD

neuron Brain cells that store and transmit information

primate a group of mammal species including monkeys, apes and humans

Allele one of the two copies of a gene

genome the name given to all the genes that contain the complete instructions for making a person or other organism

CRISPR A system for editing DNA in precise ways

Exons The small fraction of our DNA that is directly used to instruct cells how to make proteins

ASOs A type of gene silencing treatment in which specially designed DNA molecules are used to switch off a gene

RNA the chemical, similar to DNA, that makes up the 'message' molecules that cells use as working copies of genes, when manufacturing proteins.

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